



GENERAL DISCUSSION ABOUT PTS 2019 AND 2020

Helena Höök and Joakim Skarin

EURL-Campylobacter workshop 2019

FINAL RESULTS

- Final results (which are used for calculation of performance) asked for first in the Questback survey
- Please check your results as they are e-mailed to you
- You can change your replies until last day of participation
- No results will be changed after the last day of participation
- Please check preliminary report to make sure we have not made mistakes

CHANGES PTS 2018 AND 2019 AS COMPARED TO BEFORE

- Allowed for participants to select **methodology** (although ISO 10272 was recommended)
- PT 'enumeration' was mandatory (due to introduction of the process hygiene criterion) but PT 'detection' was **voluntary** (due to repeatedly good results)
- **Freeze-dried *Campylobacter*** was used for PT 'detection' in order to get more stable samples (will also allow to get closer to the detection limit)
- Introduction of **educational samples**

CHANGES PTS 2019 AS COMPARED TO 2018

- Did not offer detection as an voluntary part of PT 'enumeration' (food matrix), but allowed participants to **choose** between food and primary production **matrix** in PT 'detection'
- Provided 10 samples (instead of 18) for the PT 'detection'
- Introduction of **subtyping PT**

SOME OTHER THOUGHTS

- Recommendation on method and **procedure**
- The overall **scoring** system

PROFICIENCY TEST NO. 26

- Enumeration (and voluntary species identification) of *Campylobacter* in **chicken skin**
- The matrix will be naturally contaminated (non-sterilized)
- *Campylobacter* and other bacteria relevant to the matrix will be provided in **freeze-dried** format to be inoculated by the participants
- Participation will be **mandatory**

PROFICIENCY TEST NO. 27

- Detection and species identification of *Campylobacter* in **caecal content**
- The matrix will be produced to mimic naturally contaminated caeca (non-sterilized), but adapted to allow distribution of a homogenised sample that can be mixed with freeze-dried bacteria by the participant
- *Campylobacter* and other bacteria relevant to the matrix will be provided in **freeze-dried** format to be inoculated by the participants
- Participation will be **voluntary**
- No educational samples this year

PROFICIENCY TEST NO. 28

A PT FOR WGS QUALITY

- Objective of PT 28 is to evaluate sequence quality – from already prepared DNA and starting from bacteria
- Participation is not mandatory
- PT 28 will include 2 samples of purified DNA of *Campylobacter jejuni* and the 2 strains from which the DNA originated from
- Extract DNA for a total of 4 samples to be sequenced
- Methodology to be used: DNA-extraction + WGS
- Data to submit: information about methods and the sequence raw data

PT 28: A PT FOR WGS QUALITY

- Deadline – before summer. End of May?
- Vials with the strains?
- After cultivation and DNA-extraction, prepare sequencing libraries as soon as possible to eliminate possible stability issues of the extracted DNA.
- You do not have to perform any analysis steps or interpretations – just upload the raw data.
- Less is more? – The MiniSeq-lab got excellent assemblies and 99% of alleles were called in PT 25 for ~35X coverage and 2 x 150 bp reads. So please don't send us a NovaSeq lane of data. The server will not accept that amount of data either.
- EURL will perform different analysis of the data comparing the data with the complete genomes of the two strains.
- Gene targets such as the ones used in MLST and AMR-genes etc. might be used to test the useability of the data.

TIME PLAN FOR 2020



- All tests (PT 26–PT 28) will be sent at the same time
- Date for dispatch 9th of March?
- Survey PT 26 and PT 27 close at about 15th of April
- Survey PT 28 (subtyping) closes at about 1st of June